

Review

Animal models of cardiac arrhythmias¹

Michiel J. Janse^{a,*}, Tobias Opthof^a, André G. Kléber^b

^aDepartment of Clinical and Experimental Cardiology, Academic Medical Centre, University of Amsterdam, Amsterdam, Netherlands

^bPhysiologisches Institut, Universität Bern, Bern, Switzerland

Received 8 October 1997; accepted 3 December 1997

1. Introduction

When surveying the literature with the intention of evaluating to which extent studies on animal models have contributed to the understanding of arrhythmia mechanisms in patients and in devising therapeutic strategies, one is struck by the differences between supraventricular and ventricular arrhythmias. In general, in the field of supraventricular arrhythmias there has been a strong interaction between experimental and clinical studies and there can be no doubt that the various animal models have been instrumental in understanding the mechanisms of clinical arrhythmias and in establishing different forms of therapy. Clearly, an animal cannot be transformed into a human patient, but despite species differences and differences in arrhythmogenic factors in animal models and humans, the similarity between arrhythmia mechanisms in experimental models and patients far outweigh the differences.

This similarity is less evident when considering ventricular arrhythmias. There are several reasons for this. First, many ventricular arrhythmias, such as those induced by acute ischaemia, cannot be studied in human patients because they occur unpredictably in situations where electrophysiological changes may develop within minutes. Second, even when in patients acute ischaemia is the trigger for arrhythmias, many other factors may influence arrhythmogenesis, such as the presence of a healed infarct, hypertrophy, dilatation, electrolyte disturbances or heart failure. Third, many factors determine whether, and if so, how often ventricular arrhythmias occur in the setting of acute ischaemia and/or a chronic myocardial infarction, and in experimental models usually only a single factor is taken into account. Still, the knowledge of arrhythmogenic mechanisms derived from animal studies has greatly

contributed to the development of diagnostic and therapeutic strategies.

2. Supraventricular arrhythmias

2.1. Re-entrant tachycardias in the presence of accessory atrioventricular pathways

The history of these arrhythmias is rather bizarre because animal studies provided the basic arrhythmia mechanisms long before the syndrome was clinically recognized, because from 1967 onwards clinical studies unravelled in great detail the electrophysiological characteristics in patients without the investigators being aware of the early animal studies, and because to our knowledge only one single dog had been studied that possessed an accessory atrioventricular pathway.

In 1913 Mines described an experiment on a ring-like preparation of a tortoise heart in which he was able to initiate circulating excitation by electrical stimulation. He made the historical prediction: “I venture to suggest that a circulating excitation of this type may be responsible for some cases of paroxysmal tachycardia as observed clinically” [1].

After reading Kent’s report [2] in which a human heart was described with a muscular connection between the right atrium and the right ventricle, Mines wrote in 1914: “I now repeat this suggestion in the light of the new histological demonstration by Stanley Kent that the muscular connection between auricles and ventricles is multiple. Suppose that for some reasons an impulse from the auricle reached the main A–V bundle but failed to reach this ‘right lateral’ connection. It is possible then that the ventricle would excite the ventricular end of this lateral connection,

*Corresponding author.

¹Guest Editor was Prof. W. Schaper.

Time for primary review 42 days.

not finding it refractory as normally it would at such a time. The wave spreading then to the auricle might be expected to circulate around the path indicated" [3]. This was written 16 years before Wolff, Parkinson and White described the clinical syndrome that now bears their name [4], 18 years before Holzmänn and Scherf ascribed the abnormal ECG in these patients to pre-excitation of the ventricles via an accessory atrioventricular bundle [5], and 53 years before the first studies in patients employing intraoperative mapping and programmed stimulation proved Mines' predictions to be correct [6–8]. At present, all the electrophysiological characteristics of accessory atrioventricular connections and their role in causing re-entrant tachycardias have been obtained in studies on human patients (for review see Wellens [9]) and only one study described pre-excitation in a dog [10]. In this study, which also described two patients, the important observation was made that atrial fibrillation induced in the dog, caused ventricular fibrillation as well because the accessory pathway had a short refractory period and conducted many impulses, which otherwise would have been blocked in the AV node. Still, at present there is certainly no need for an animal model of accessory atrioventricular pathways.

2.2. Atrioventricular nodal re-entrant tachycardia

The history of this arrhythmia is very different from that of the arrhythmias caused by accessory AV connections. Although it was again Mines [1,2] who formulated the basic mechanisms, clinical studies quickly followed and throughout this century there has been an intensive interaction between experimental and clinical studies (for review see [11] and [12]). For example, the first study employing programmed stimulation in patients to unravel the arrhythmia mechanisms and to treat the condition by pacemaker implantation by Coumel and co-workers in 1967 [13] quoted the early studies of Mines [1,2]. Also, the pioneering clinical studies of the 1980s allowing successful surgical treatment [14,15] or catheter ablation [16–18] of the arrhythmia, all quoted the microelectrode studies of the 1960s and 1970s on isolated rabbit heart preparations that provided insight into arrhythmia mechanisms on a cellular basis [19–22].

Today, we are confronted with a rather paradoxical situation: the history indicates a happy union between knowledge gathered by both experimental and clinical studies, the former preceding the latter, which finally resulted in the very successful treatment by radiofrequency catheter ablation [23–25]. Despite this success, there are still many uncertainties about the exact location of the re-entrant pathway and about the electrophysiological and structural properties of the two AV nodal pathways ('slow' and 'fast') that are thought to form the basis for AV nodal re-entry. The animal model most often used, the isolated superfused rabbit heart preparation, differs from the heart

of patients suffering from AV nodal re-entrant tachycardia. It is extremely rare to induce sustained AV nodal re-entry in this preparation, but it is not uncommon to induce single echo beats by premature stimulation of the atrial or ventricular tissue [19–22,26–29].

In open-chested anaesthetized dogs, only one 'case report' of sustained re-entrant AV nodal tachycardia has been published [30], and reproducibly induced sustained tachycardia became only possible after a surgical procedure that blocked atrial impulses from the anterior input site to the AV node [31].

In patients with AV nodal re-entrant tachycardia, the hallmark for dual AV nodal pathways is the so-called 'jump' in the conduction curve [32]: when during regular pacing of the atria (A1) single premature atrial stimuli (A2) at progressively shorter coupling intervals are applied and the premature atrium–His bundle intervals (A2–H2) are plotted against the A1–A2 intervals, a sudden jump of 50 ms or more in the A2–H2 interval at a decrement in A1–A2 of 10 ms is taken as evidence for dual AV nodal pathways. At a critical coupling interval the 'fast' pathway is refractory, and conduction to the His bundle proceeds via the 'slow' pathway which has a shorter refractory period. In isolated rabbit heart preparations, such a jump has not been observed [33]. In the anaesthetized dog, a 'jump' was observed in one study only after left and right stellate ganglia had been removed [34]. In another study, no jump was found [35]. In yet another study, a jump in the A–V interval was ascribed to slow conduction or block in the peripheral Purkinje system rather than to functional dissociation of the AV node [36]. In isolated, perfused dog hearts, no jump occurs [37,38], although ventricular echoes could be reproducibly induced [39]. Whereas the clinical experience would suggest that atrial tissue is involved in the re-entrant circuit (for review see [12] and [40], but for a different point of view see [41]), mapping studies in the isolated, blood perfused dog heart showed that ventricular echoes are due to subatrial re-entry [39].

It would be desirable if an animal model could be found in which subtle manipulation of the autonomic nervous system (rather than a surgical procedure as described in [31]) would result in sustained AV nodal tachycardia being reproducibly induced. Answers could then be found to questions such as: Is the re-entrant circuit for single echo beats the same as that for sustained re-entry? Are there structurally different pathways? Are there multiple re-entrant circuits? What is the role of anisotropic conduction and of different input sites? In any case, there is no animal model for dual pathways as observed in humans.

2.3. Atrial flutter

Much of our knowledge about activation patterns during atrial flutter has been derived from animal studies, with the 1920 paper of Lewis, Feil and Stroud as the classical example [42]. Catheter electrode mapping in patients was

first performed by Puech and colleagues in 1956 [43] and due to advances in mapping techniques, recent studies in man [44] allowed comparison to data obtained in animal models [45]. On the basis of these studies, it is widely accepted that atrial flutter is due to re-entry.

There are a number of animal models in which atrial flutter was induced following the creation of anatomical lesions, by extending an anatomical obstacle such as the ostium of the vena cava superior by crushing the intercaval region [46,47], or by producing single lesions in the right atrium [48–50]. It is doubtful whether these models are representative of atrial flutter in patients.

Another type of flutter, depending on a Y-shaped surgical lesion in the right atrial free wall, causing the re-entrant circuit to consist of atrial tissue around the tricuspid ring [51] may have a clinical counterpart in postoperative flutter following surgical correction of congenital abnormalities [52].

The canine model developed by Boyden and Hoffman [53], in which right atrial enlargement was produced by banding of the pulmonary artery and by producing tricuspid regurgitation, may also have a clinical counterpart in patients with chronic obstructive pulmonary disease and tricuspid regurgitation. In those dogs, a functional zone of block and area of slow conduction set the stage for re-entry, rather than an anatomical obstacle. Functional re-entry is also observed in the sterile pericarditis model of canine atrial flutter, first described by Pagé et al. [54]. This model was developed because of the fact that following cardiac surgery in patients atrial flutter frequently occurs and this may be related to postoperative sterile pericarditis. Another form of functional re-entry causing atrial flutter was pharmacologically induced in isolated dog hearts by infusion of acetylcholine [55].

Despite subtle differences in the various forms of atrial flutter, one cannot but agree with Mary-Rabine and co-workers [56], that “the history of atrial flutter clearly illustrates the bidirectional flow of information and the mutual stimulation between the basic and the clinical levels, leading both to a better understanding of the nature of the arrhythmia and to new therapeutic approaches”. It is now established that atrial flutter is due to a re-entrant wave in the right atrium, and that a zone of slow conduction located inferiorly and posteriorly in the right atrium is the target for catheter ablation.

2.4. Atrial fibrillation

Some crucial observations on the characteristics of atrial fibrillation were made long before technical developments allowed the simultaneous recording from multiple atrial sites, a prerequisite for documenting the complex activation sequence of fibrillation. Thus, in 1914 Garrey [57] without the help of electrophysiological or mechanical recordings established that a critical mass of tissue is needed to sustain fibrillation. He also suggested that

fibrillation was due to “... a series of ring-like circuits of shifting location and multiple complexity” [58]. Some 70 years later, Allesie and co-workers could record simultaneously from 192 atrial sites in an isolated, Langendorff-perfused canine heart in which atrial fibrillation was induced by rapid atrial pacing during infusion of acetylcholine [59]. In essence, they confirmed Garrey’s statement, which had been refined by Moe and Abildskov on the basis of both experimental observations and computer simulations that led to the formulation of the multiple wavelet hypothesis [60,61]. In the experiments of Allesie et al. [59] the presence of multiple independent wavelets was documented, and it was estimated that the critical number of wavelets in both atria necessary to maintain fibrillation was between 3 and 6. These results were largely confirmed by other studies in dog hearts [62] and also by mapping studies in patients with atrial fibrillation during open heart surgery [63,64].

In recent years, there has been a remarkable interaction between experimental and clinical studies, where important findings in animal models were soon thereafter confirmed in clinical studies. For example, repetitive induction of atrial fibrillation, or prolonged periods of rapid atrial pacing, in conscious goats gave rise to a marked shortening of the atrial refractory period, which persisted for a long time after restoration of sinus rhythm and predisposed to the reinduction of atrial fibrillation following cardioversion [65]. This was confirmed in patients [66–68], as was the finding originally made in canine hearts [70], that calcium blocking agents could attenuate this shortening of the refractory period [69]. Despite the great impact that the experimental studies have on clinical developments, including new strategies for therapy such as catheter ablation (see Ref. [67]), there are obvious differences between the atria of the experimental animals used and those of patients who spontaneously develop atrial fibrillation. In the experimental models, atrial fibrillation was induced in essentially normal hearts, either by shorter or longer periods of rapid pacing or repetitive induction of atrial fibrillation by burst pacing [65,70] or by pharmacologic means, such as infusion of acetylcholine [59,62]. Whilst this latter model may have its clinical counterpart in the relatively rare form of atrial fibrillation dependent on increased vagal tone [71], about 85% of patients with atrial fibrillation have an underlying structural cardiac abnormality or a metabolic disorder, often associated with atrial enlargement [72]. Acute atrial dilatation shortens the atrial refractory period and enhances the vulnerability to atrial fibrillation [73,74]. However, the effects of chronic stretch are most likely different from those of acute stretch, and would be more important in contributing to atrial fibrillation in patients. There is one study by Boyden et al. [75] in which atrial cellular electrophysiological characteristics of dogs with naturally occurring mitral valve disease leading to progressive atrial enlargement were studied. Some animals were followed for 5 years, before the electrophysiological study

was performed. Most dogs developed atrial arrhythmias, including atrial fibrillation. Surprisingly, the transmembrane potential characteristics of atrial cells of these animals were not significantly different from those of control animals, although some cells were found with resting membrane potentials below -60 mV that were inexcitable. This is in contrast to the findings of the studies mentioned earlier [65,66] and also in contrast to several studies in which cellular electrophysiological abnormalities have been documented in small, isolated atrial preparations obtained from fibrillating human atria [76–78]. In the study of Boyden et al. [75], massive interstitial fibrosis and cellular hypertrophy were found and the authors concluded that the morphological changes were much more important in causing atrial fibrillation than the slight, insignificant electrophysiological alterations they found. The increased size of the atria would permit the coexistence of many re-entrant circuits. The increase in connective tissue would promote inhomogeneous conduction, unidirectional block and re-entry [79]. Thus far, the emphasis in animal models of atrial fibrillation has been on electrophysiological characteristics in structurally normal hearts. To further diminish the gap between animal models and patients, the arrhythmogenic effects of structural changes deserve further study.

3. Ventricular arrhythmias

3.1. A hereditary model of sudden death

A colony of German shepherd dogs has been described with inherited ventricular arrhythmias and a predisposition for sudden death [80]. Sudden death most often occurs during sleep or at rest after exercise or excitement. The electrocardiogram does not show a prolonged QT interval, but frequently there is marked notching of the T wave. The arrhythmias are rapid polymorphic ventricular tachycardias, following long R–R intervals, and are most likely due to triggered activity induced by early afterdepolarizations in the Purkinje system [81]. In epicardial myocytes, the density of the transient outward current (I_{to}) was reduced, and the time constant of inactivation was reduced [82]. In addition, deficiencies in cardiac sympathetic denervations have been reported [83]. At first glance, this dog model bears a resemblance to the congenital long QT syndrome in which bradycardia induced polymorphic ventricular tachycardia and sudden death occur and in which genetic defects in ion channels regulating repolarization have been described [84,85]. However, the dogs have no prolonged QT interval and thus far, in patients with the long QT syndrome no deficiencies in I_{to} have been described [85]. Still, this animal model might have a clinical counterpart because patients have been described with polymorphous ventricular tachycardia (Torsade de pointes) who have a normal QT interval [86,87].

3.2. Ventricular arrhythmias caused by acute ischaemia

A great many experimental studies on this subject have been undertaken in the past decades. A rough distinction can be made into studies in which electrophysiological parameters relevant for understanding arrhythmia mechanisms were recorded (for Refs. see [88,89]), and those in which only the incidence of the lethal arrhythmia, ventricular fibrillation, was noted, usually in studies testing anti-arrhythmic drugs.

Validation of animal models for assessing the pathophysiology of acute myocardial ischaemia implicates consideration of the (1) diversity of cardiac diseases which involve acute ischaemia and (2) the variety of experimental models which have mostly been designed to mimic part of the complex events occurring in the human disease. Along this line of reasoning, mostly large animals (dogs, pigs, cats) are used to study ventricular arrhythmias, while mostly small animals (rats) are involved in studies about the changes in metabolic pathways consequent to ischaemia and reperfusion. In the former, multisite mapping of electrical activity is applied to the analysis of the mechanisms of ventricular tachycardia and fibrillation [90,91] whereas in small animals' hearts, which primarily serve the purpose to provide numerous and affordable samples for chemical analysis, arrhythmias are defined by ECG patterns and analyzed statistically on the basis of their incidence. Thus the main reason for selecting a certain animal species appears to be the suitability for the application of a specific technique, assuming that 'acute' human ischaemia/reperfusion can be compared with experimental ischaemia/reperfusion independently of the species selected. While this assumption might hold for the very global and basic changes in metabolism, it is doubtful that such a simplified concept is applicable to the study of arrhythmogenesis, as outlined below. According to a search in 'MEDLINE' a total of 1327 studies have been carried out between 1966 and 1996 for the assessment of ventricular fibrillation in the setting of myocardial ischaemia. 569 of these studies were carried out in dogs, 126 in pigs, 25 in guinea pigs, 51 in rabbits and 931 in rats. In the setting of reperfusion arrhythmias, a total of 1159 studies were carried out (328 in dogs, 128 in pigs, 54 in rabbits, 64 in guinea pigs, 585 in rats).

The fact that acute ischaemia affects the incidence of arrhythmias in coronary heart disease in a variety of ways makes it impossible to investigate its pathophysiology in a single experimental model. Thus, acute ischaemia occurs as one of the triggers for arrhythmias in chronic infarction [92], in hypertrophy and failure, or it may occur consequently to coronary occlusion (thrombosis or spasm) in previously relatively healthy individuals [93]. This spectrum of pre-existing alterations is likely to affect the role of acute ischaemia, since infarction, hypertrophy and failure are associated with changes in the pre-existing arrhythmogenic substrate [94,95]. Moreover, experimental-

ists distinguish between total and partial coronary occlusion, the latter being associated with so-called 'low flow' ischaemia, a pathophysiologic entity which leads to electrical and ionic changes different from the changes associated with immediate and total occlusion [96,97]. In the clinical settings, it is not always evident whether acute myocardial ischaemia is associated with or without residual flow through the occluded or collateral arteries. In the light of these complexities the discussion about the applicability of animal models of acute ischaemia to the human situation is certainly justified.

A first basic question concerns the comparison of the type and incidence of the electrical changes and arrhythmias among different species. In larger species (dog, pig, cat) the arrhythmias during acute ischaemia are relatively well characterized. Thus arrhythmias occur in two distinct phases (so-called 'phase IA' and 'phase IB', Ref. [98]). These two phases are associated with distinct changes in the electrical tissue properties. In the first phase, IA, (up to approximately 8–10 min of coronary occlusion), there is a rapid change in electrical membrane properties associated with metabolic acidification (anaerobic glycolysis), and cellular loss and extracellular accumulation of $[K^+]_o$ [99,100]. The impact of these changes are a rapid depolarization of the ischaemic myocytes, and a loss of amplitude and duration of the transmembrane action potential [101,102]. Moreover, there is a marked lengthening of the refractory period, which becomes sensitive to the lengths of the previous intervals of local excitation. As shown in experimental studies and explained recently in computer simulations [103,104], these changes are followed by specific changes in the excitation and conduction patterns. Thus, the decrease in conduction velocity during acute ischaemia is relatively small and conduction block, which changes its location from beat-to-beat, occurs early and abruptly [104]. The resulting ventricular tachycardia, frequently issuing into fibrillation, has a characteristic appearance [90]. It is made up by large and highly unstable re-entrant circuits (several millimetres inner circle length). The second phase of arrhythmias, IB, occurs approximately 10 to 15 min after coronary occlusion and is related to the electrical uncoupling of the myocytes [105]. Although the IB arrhythmias have not been analyzed in detail by mapping studies, it is likely that the re-entrant circuits during this phase are significantly smaller, because partial electrical uncoupling allows for much smaller conduction velocities [106], and therefore scales the circus movements to a smaller size. The studies obtained in relatively large animals (dog, pig, cat) raise two questions: (1) Are the arrhythmias occurring after acute coronary occlusion in humans comparable to those observed in the animal models? (2) Do the experimental arrhythmias differ among animal species? The first question cannot be answered directly. However, several observations provide circumstantial evidence that the electrical changes during acute ischaemia in humans and large animal hearts are similar. In

an experiment carried out in an isolated perfused human heart, the extent and time course of the changes in transmembrane action potentials were almost identical to those observed in pig and dog hearts [107]. While there seems an approximate similarity among the electrophysiological changes observed in large animals and in certain cases of acute ischaemia in humans, several experimental observations indicate that the arrhythmias observed in small animals (guinea pigs, rats) during acute ischaemia differ from the arrhythmias observed in larger species. First, as mentioned above, the very early (and frequent) type IA arrhythmias require a large tissue mass for the re-entrant circuits to be maintained. The dependence of re-entry (tachycardia and fibrillation) on tissue size is an old observation, made already in 1914 by Garrey [57]. Indeed, most of the work done on ventricular fibrillation in rat hearts indicates that VF in this species occurs at a time corresponding to the IB arrhythmias, and no clear separation of IA from IB arrhythmias has been described in small animal species.

A number of further processes critical for the electrical changes in acute myocardial ischaemia might be different in rats and guinea pigs from larger species. Extracellular accumulation of $[K^+]_o$ and metabolic acidification, which are likely to be linked to each other, are the main factors determining the extent of the depolarization of the resting membrane, the changes in action potential upstroke and changes in refractoriness. In ischaemic regions devoid of collateral flow, there is a sharp transition from ischaemic to normoxic tissue with respect to local pO_2 . In contrast, the acidification of the tissue, and the extracellular accumulation of $[K^+]_o$ show a gradual decrease from the centre towards the border of the ischaemic region [108,109]. In other words, although a given tissue site may be fully ischaemic, the extent of the ionic changes, which form the basis for the disturbance in electrical function, depends on the diffusion of products of the ischaemic metabolism toward the non-ischaemic border. Both K^+ and CO_2 have been invoked in this diffusion process [110,111], whereby CO_2 seems to be a particularly important factor, because it has a high diffusion coefficient and is bound in large quantities by the carbonic buffer system [112,113]. Along this line of reasoning, it has to be assumed that the process of diffusion will not only affect the gradients between the centre and the border of an ischaemic region, but it will also lead to a relatively smaller change in extra- and intracellular pH and K^+ in hearts with a small ventricular mass, and consequently affect the electrical changes of the ischaemic myocytes [111]. In a series of experiments in which extracellular potassium $[K^+]_o$ accumulation was compared among a variety of species, the maximal $[K^+]_o$ levels reached during acute ischaemia in rats were by approximately 20–30% lower than those observed in guinea pigs or rabbits [113,114].

Besides the macroscopic dimension of the ventricles, intrinsic differences in normoxic metabolism may also

exist among species. In rats such differences have been shown with respect to the duration and shape of the transmembrane action potential and the homeostasis of cellular Na^+ and Ca^{2+} . Thus the relatively high normal intracellular Na^+ activity in the normal rat affects the working mode of the Na/Ca exchanger and may lead to earlier Ca^{2+} overload in depolarized cells. Interestingly in rats, this difference can be reversed by inhibition of thyroid hormone production [115].

In the past years, so-called remodelling of cardiac tissue in various cardiac diseases and with a variety of stimuli has gained a wide interest. Many of these remodelling processes which occur in ischaemia, chronic infarction, myopathy, hypertrophy and failure, may change the substrate for electrical excitation and conduction at any level. Thus, the macroscopic tissue architecture may get more discontinuous via the increase in connective tissue. The expression of gap junctions can change as well, as may the expression of a large number of membrane ionic channels responsible for excitation. The results of all these studies demonstrate that assessment of pathophysiological mechanism has to consider the dynamics of events and related variables not only on short term (e.g. in the minutes following coronary occlusion) but also on middle (e.g. after preconditioning [116,117]) and on longer term (genetic remodelling [118–121]). This increasing complexity should have an impact on the selection and definition of animal models. First, in many important diseases, it is probably wrong to think that a given animal model can exactly mimic a certain disease and the ‘individuality’ of a certain disease pattern. In most cases they can only address partial aspects of the disease mechanism. Second, animal models used to assess arrhythmias should be designed in such a way that the most important variables methodologically followed.

The factors that determine whether, and if so, how frequently, ventricular fibrillation occurs include the size of the ischaemic area, the degree of collateral flow, heart rate, the use of anaesthetics, stress in conscious animals, the mode of coronary artery occlusion, presence of a previous infarction, activity of the autonomous nervous system, hypertrophy in the non-ischaemic myocardium [88,89]. The three most important factors are size of the ischaemic area, the degree of collateral flow and heart rate [122–127]. There is a variation among species in the degree of collateral blood flow following coronary artery occlusion [96]. For example, in rat, rabbit and pig hearts collateral flow is not significantly different from zero, in the guinea pig it is not different from normal control flow [128]. In the dog, an animal often used in studies on ischaemia-induced arrhythmias, there is a variation in pre-existing collaterals, and depending on the degree of collateral flow, the incidence of ventricular fibrillation may vary from zero to 100% after occlusion of a major coronary artery [124,126,127]. Similarly, occlusion of the left anterior descending coronary artery results in a vari-

ation of size of the ischaemic zone [122–124] and “this can account for a substantial portion of non-drug related variability in outcome of antiarrhythmic trials using the canine coronary occlusion or release model” [123]. This statement was corroborated by Trolese-Mongheal and colleagues [129] who collected data from various laboratories on 658 dogs in which the left anterior descending coronary artery was suddenly ligated. When control series consisted of 10 dogs, the incidence of ventricular fibrillation varied from 0 to 70%; when the control group counted 20 animals, the incidence varied from 5 to 55%, and even in series of 100 dogs, there still was a range of 14 to 36%. The papers emphasizing the importance of pre-existing collaterals were published between 1970 and 1986. A Medline search unearthed 28 studies published between 1982 and 1996 in which a major coronary artery was occluded in dogs to test the effect of antiarrhythmic drugs. The control series varied from 6 to 40 animals, and since the factors mentioned above were not controlled, interpretation of the results must be made with great caution.

3.3. The ventricular arrhythmias of myocardial infarction

A distinction has been made in arrhythmias occurring in the subacute phase of myocardial infarction (hours to days after acute obstruction of a coronary artery) and in the chronic phase (weeks to months). Almost all of the experimental work has been performed in dogs (a Medline search over the past 4.5 years identified 25 dog studies, one in the rat and one in the pig).

The spontaneous arrhythmias that occur in the dog during the subacute phase resemble those in patients recovering from an acute myocardial infarction: accelerated idioventricular rhythms or slow ventricular tachycardias that usually do not degenerate into ventricular fibrillation (for a detailed description see Refs. [88] and [89]). A great deal is known about electrophysiological changes in both Purkinje and muscle cells that survive the infarct. However, since the arrhythmias of the subacute phase are usually benign, and since the patients are still in hospital so that in case ventricular fibrillation would occur adequate resuscitation and defibrillation will be provided, the knowledge derived from experimental models has not contributed to establish therapeutic strategies during this phase of myocardial infarction.

A great deal of information about the electrophysiological characteristics of ventricular tachycardia induced by programmed electrical stimulation has been gathered in dogs with a healing infarct, and many similarities exist between these arrhythmias and those induced in patients. There are, however, several differences between the experimental and the clinical tachycardias. Thus, in dogs, ventricular tachycardias can be easily induced by premature ventricular stimulation in the first week following coronary artery occlusion, but after the first week in-

ducibility decreases [130] and sometimes the arrhythmia cannot be induced at all [131]. This is different in human patients: after five days, ventricular tachycardias can be induced in about 10% of patients [132], but in 20 to 50% after three weeks [132–134]. Moreover, in the dog, the re-entrant circuit responsible for the tachycardia is usually located in the so-called subepicardial border zone, i.e. the thin layer of surviving subepicardial myocardium overlying the infarct, whereas many of the sustained tachycardias in humans with a healed infarct originate in the subendocardial region [89]. Despite these differences, the experimental studies have provided important information about the characteristics of the re-entrant circuits and in so far studies in humans were able to study these characteristics, the similarities far outweigh the differences. There is no doubt that the experimental findings have been of crucial importance for initiating therapeutic strategies, such as mapping-guided surgery, catheter ablation, or antitachycardia pacing [135–137]. The problems arise when experimental studies are performed without electrophysiological measurements, noting only the incidence of arrhythmias. As is the case for acute ischaemia, many factors determine the incidence of ventricular tachycardia or fibrillation in a heart with a healed infarct. Both the size and the structure of the infarct determine whether or not arrhythmias occur and the characteristics of arrhythmias that do occur [138,139]. Without the presence of surviving muscle fibres in the infarcted region that provide the anatomical substrate for re-entry, arrhythmias may not occur [140–142]. The location of the surviving myocardial fibres might also determine whether the autonomic nervous system might contribute to arrhythmogenesis. Since efferent sympathetic fibres travel in the left ventricular subepicardium, a transmural infarct extending to the epicardial surface may damage them and produce nonhomogeneous sympathetic denervation of normal myocardium distal to the infarct [143], which may be arrhythmogenic [144,145]. These facts are important for the interpretation of animal experiments in which only one factor important for arrhythmogenesis is considered. The studies of Schwartz and colleagues have been instrumental in initiating clinical studies on baroreflex sensitivity as a risk factor for arrhythmias in post-infarction patients [146–148]. In essence, their experimental model is a conscious dog with a healed anterior infarct in which during exercise, an occluder on the circumflex coronary artery is occluded for two minutes. It appeared that dogs with low baroreflex slopes (susceptible dogs) developed more often ventricular fibrillation than dogs with steep baroreflex slopes (resistant dogs). These results were interpreted as indicating that strong vagal reflexes would protect an individual with an infarction against stress- and ischaemia-induced ventricular fibrillation. Indeed, clinical studies [149,150] have confirmed that baroreflex sensitivity is an important determinant for sudden death and inducibility of ventricular tachycardia in post-infarction patients. Still, both in ani-

mals and patients there is a considerable overlap in baroreflex slope in the groups with and without arrhythmias, so that in an individual case no absolute prediction can be made whether or not arrhythmias will develop. This is of course due to the fact that sudden death is not due to a single pathophysiologic event. It is in this respect noteworthy that Legato found that susceptible dogs had larger and more inhomogeneous infarcts than resistant dogs [151].

3.4. Animal models and antiarrhythmic drugs

Generally speaking, antiarrhythmic drugs exert their effects largely by modulating conduction velocity, or refractory period duration, or both. Conduction velocity depends on the one hand on the passive electrical properties of cardiac tissue, on the other hand on the characteristics of the Na^+ channels and Ca^{2+} channels. Whilst there is some evidence that, at least in the Purkinje system, conduction velocity increases in proportion to the size of the heart, most likely due to an increase in the space constant [152,153], to our knowledge very little is known about species differences in the density and kinetics of Na^+ and Ca^{2+} channels. In contrast, there are marked differences among species in the K^+ currents that largely determine repolarization, so that action potential duration and duration of the refractory period differ widely in various species. For that reason, we will concentrate on species differences in refractory period duration.

It is generally assumed that agents that prolong the action potential duration, and thereby the refractory period, are effective against re-entrant arrhythmias in two ways: by prolonging the wavelength (the product of refractory period and conduction velocity), the initiation of a re-entrant arrhythmia by a premature impulse may be prevented [154] or an existing arrhythmia may terminate because the wavelength becomes too large with respect to the re-entrant circuit, so that by closing the excitable gap, the head of the re-entrant wavefront will hit the wall of refractoriness and propagation stops [155a,b]. Even though these explanations may be too simplistic because some studies have shown that re-entrant arrhythmias may be terminated by agents that prolong the refractory period without entirely closing the excitable gap [156], it is clear that the duration of the diastolic interval is an important parameter when assessing the efficacy of an anti-arrhythmic drug that prolongs refractoriness in an animal model.

Fig. 1 shows the relation between the basic cycle length and the action potential duration or effective refractory period in the ventricle (or in isolated trabeculae or isolated cells) in several species [157–161]. In contrast to *all* other species it may be appreciated that in the rat there is no shortening of refractory periods at the shorter cycle lengths. Also, there are considerable differences in refractory periods between the dog and the pig, although

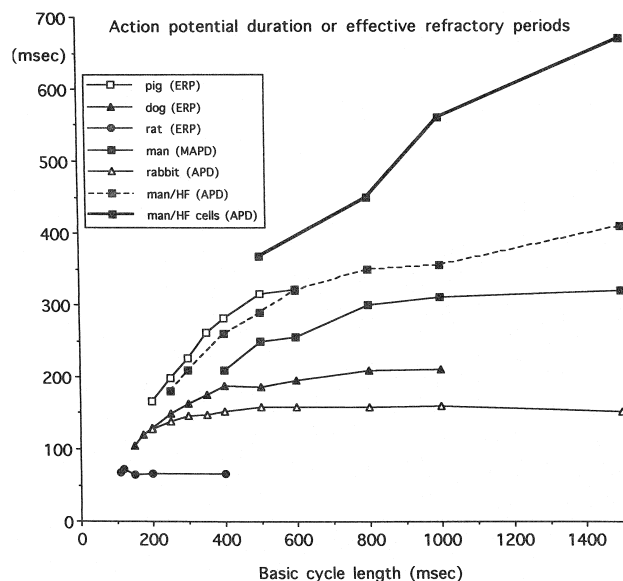


Fig. 1. Relation between (steady state) cycle length and (monophasic) action potential duration or effective refractory period in several species (see inset for details). Data have been taken as follows: pig [159], dog [157], rat [158], man [161], rabbit [160], man/HF (heart failure) [160], man/HF (heart failure isolated cells (unpublished data, Veldkamp MW)). ERP: effective refractory period; APD: action potential duration; MAPD: monophasic action potential duration.

these species have comparable cardiac dimensions and in vivo and in vitro cycle lengths. The discrepancy between the monophasic action potential duration in the normal human ventricle (Fig. 1: filled squares, solid line) and action potential duration in ventricular trabeculae from failing human hearts (Fig. 1: filled squares, dashed line) underscores the prolongation of action potential duration associated with heart failure. In the latter study [160] patients were selected who were not on amiodarone, quinidine or sotalol, thereby excluding drug effects in addition to the effect of heart failure itself. Action potential duration in single cells isolated from failing human hearts (Fig. 1: filled squares, fat line) is even further prolonged compared to trabeculae. It cannot be determined with certainty whether this follows from the cell isolation procedure, but it certainly sets limits to the choice of a *relevant animal model*. Fig. 2 shows the diastolic interval along the ordinate (log scale used for better description of the data at the more relevant shorter cycle lengths) versus the cycle length. Fig. 2 shows that rat ventricle is not the first choice if one aims at ‘filling up’ the diastolic interval by means of a class I or class III or an experimental anti-arrhythmic agent. On the other hand, porcine ventricular myocardium appears to have a diastolic interval similar to that in human ventricle. In line with these data it may be interesting to assess the number of studies performed in several species on the effects of anti-arrhythmic agents on action potential duration. Table 1 shows the results of a

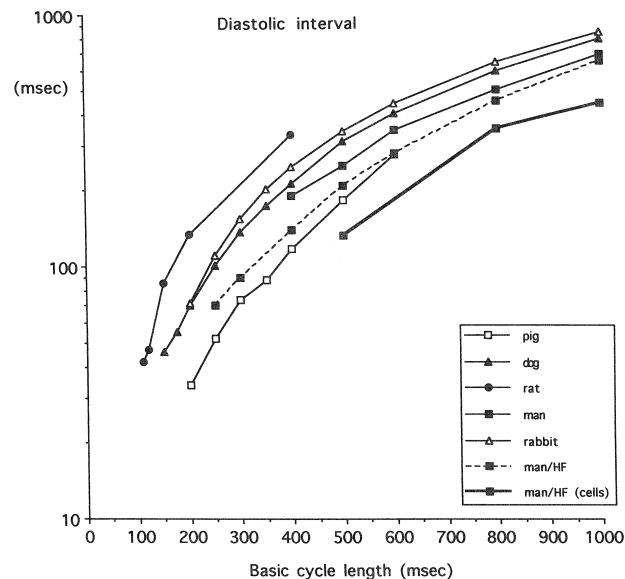


Fig. 2. Relation between (steady state) cycle length and diastolic interval (see inset for details). Data have been taken as indicated in the legend to Fig. 1.

Medline search for the number of studies performed on the effects of anti-arrhythmic agents on action potential duration in different species between 1966 and 1992, and between 1993 and 1996. The rabbit is often used for electrophysiological research, probably because it constitutes a reasonable compromise in terms of cardiac dimensions, basic electrophysiological characteristics and cost. Therefore it has been chosen as the reference species for a comparison of the species chosen in this area of research in the past and in more recent years. Table 1 shows that the use of the pig and the guinea pig has increased during recent years and that the use of the rat is small compared to other species which seems justified on the basis of data in Figs. 1 and 2. Also, the use of human tissue has increased.

Table 1
Studies on action potential duration and antiarrhythmic agents

Species	1966–1992	Relative number	1993–1996	Relative number
Dog	123	1.9	21	1.2
Pig	101	1.6	48	2.8
Cat	7	0.1	6	0.4
Rabbit	64	1.0	17	1.0
Guinea pig	100	1.6	45	2.6
Rat	19	0.3	6	0.4
Man	22	0.3	22	1.3

Absolute and relative numbers of studies performed on the effect of anti-arrhythmic agents on action potential duration in several species between 1966 and 1992, and between 1993 and 1996, as included in Medline.

4. Conclusions

It is clear that species differences do exist with respect to factors that determine arrhythmogenesis and it is also clear that no animal model will accurately mimic the human patient suffering from, or threatened by an arrhythmia.

Nevertheless, the knowledge gathered from animal studies undoubtedly has been instrumental in devising diagnostic and therapeutic strategies both in supraventricular and ventricular arrhythmias. It is our conviction that in the future, new knowledge will be obtained from experiments performed at many levels: in systems expressing and testing the functions of molecules involved in electrical excitation, in single cells, cell cultures, excised cardiac preparations, isolated whole hearts, whole hearts in anaesthetized animals, and in conscious animals. It will be the combination of such investigations, rather than a single model or experimental technique, which will lead to novel strategies for diagnosis and treatment. Finally, electrophysiological studies should be encouraged in animals with ‘naturally’ occurring cardiovascular disease [162,163].

Acknowledgements

A.G.K. was supported by the Scientific Durrer Foundation, Utrecht, Netherlands

References

- [1] Mines GR. On dynamic equilibrium in the heart. *J Physiol* 1913;46:349–383.
- [2] Kent AFS. Observations on the auriculo-ventricular junction of the mammalian heart. *Quart J Exp Physiol* 1913;7:193–195.
- [3] Mines GR. On circulation excitations in heart muscles and their possible relation to tachycardia and fibrillation. *Trans Roy Soc of Canada* 1914;section IV:42–53.
- [4] Wolff L, Parkinson J, White PD. Bundle-branch block with short P-R interval in healthy young people prone to paroxysmal tachycardia. *Am Heart J* 1930;5:685–704.
- [5] Holzmänn M, Scherf D. Ueber Elektrokardiogramme mit verkürzter Vorhof–Kammer-Distanz und positiven P-Zacken. *Z Klin Med* 1932;121:404–423.
- [6] Durrer D, Roos JR. Epicardial excitation of the ventricles in a patient with a Wolff–Parkinson–White syndrome (type B). *Circulation* 1967;35:15–21.
- [7] Burchell HB, Frye RB, Anderson MW, McGoon DC. Atrioventricular and ventriculo-atrial excitation in Wolff–Parkinson–White syndrome (type B). *Circulation* 1967;36:663–672.
- [8] Durrer D, Schöo L, Schuilenburg RM, Wellens HJJ. The role of premature beats in the initiation and termination of supraventricular tachycardia in the Wolff–Parkinson–White syndrome. *Circulation* 1967;36:644–662.
- [9] Wellens HJJ. The electrophysiological properties of the accessory pathway in the Wolff–Parkinson–White syndrome. In: Wellens HJJ, Lie KI, Janse MJ, editors. *The Conduction System of the Heart*. Leiden: Stenfert Kroese, 1976, 1988:567–587.
- [10] Boineau JP, Moore EN. Evidence for propagation of activation across an accessory atrioventricular connection in types A and B preexcitation. *Circulation* 1970;41:375–397.
- [11] Janse MJ, Anderson RH, McGuire MA, Ho SY. ‘AV nodal’ reentry: part I. AV nodal reentry revisited. *J Cardiovasc Electrophysiol* 1993;4:561–572.
- [12] McGuire MA, Janse MJ, Ross DL. ‘AV nodal’ reentry: part II. AV nodal, AV junctional, atrionodal reentry?. *J Cardiovasc Electrophysiol* 1993;4:573–586.
- [13] Coumel P, Cabrol C, Fabiato A, Gourgon R, Slama R. Tachycardie permanente par rythme réciproque. *Arch Mal Coeur* 1967;60:1830–1864.
- [14] Ross DL, Johnson DC, Denniss R, Cooper MJ, Richards DA, Usher JB. Curative surgery for atrioventricular junctional (‘AV nodal’) reentrant tachycardia. *J Am Coll Cardiol* 1985;6:1383–1392.
- [15] Cox JL, Holman WL, Cain ME. Cryosurgical treatment of atrioventricular node reentrant tachycardia. *Circulation* 1987;76:1329–1376.
- [16] Sung RJ, Waxman HL, Saksena S, Juma Z. Sequence of retrograde atrial activation in patients with dual atrioventricular nodal pathways. *Circulation* 1981;64:1059–1067.
- [17] Haissaguerre M, Warin JF, Lemetayer P, Saoudi N, Guillem JP, Blanchot P. Closed-chest ablation of retrograde conduction in patients with atrioventricular nodal reentrant tachycardia. *N Engl J Med* 1989;320:426–433.
- [18] Epstein LM, Scheinman MM, Langberg JL, Chilson D, Goldberg HR, Griffin JC. Percutaneous catheter modification of the atrioventricular node. A potential cure for atrioventricular nodal reentrant tachycardia. *Circulation* 1989;80:757–768.
- [19] Mendez C, Moe GK. Demonstration of a dual AV nodal conduction system in the isolated rabbit heart. *Circ Res* 1966;19:378–393.
- [20] Janse MJ, van Capelle FJL, Freud GE, Durrer D. Circus movement within the A–V node as a basis for supraventricular tachycardia as shown by multiple microelectrode recording in the isolated rabbit heart. *Circ Res* 1971;28:403–414.
- [21] Wit AL, Goldreyer BN, Damato AN. An in vitro model of paroxysmal supraventricular tachycardia. *Circulation* 1971;43:862–875.
- [22] Janse MJ, Van Capelle FJL, Anderson RH, Touboul P, Billette J. Electrophysiology and structure of the atrioventricular node of the isolated rabbit heart. In: Wellens HJJ, Lie KI, Janse MJ editors. *The Conduction System of the Heart*. Leiden: Stenfert Kroese, 1976, 1988:296–315.
- [23] Jackman WM, Beckman KJ, McClelland JH, et al. Treatment of supraventricular tachycardia due to atrioventricular nodal reentry by radiofrequency catheter ablation of slow-pathway conduction. *N Engl J Med* 1992;327:313–318.
- [24] Haissaguerre M, Gaita F, Fisher B, et al. Elimination of atrioventricular nodal reentrant tachycardia using discrete slow potentials to guide application of radiofrequency energy. *Circulation* 1992;85:2162–2175.
- [25] Sanjuan R, Morell S, García Civera R, et al. Transvenous ablation with high frequency energy for atrioventricular junctional (AV nodal) reentrant tachycardia. *PACE* 1989;12:1631–1639.
- [26] Iinuma HL, Dreifus LS, Mazgalev T, Price R, Michelson EL. Role of the perinodal region in atrioventricular nodal reentry: evidence in an isolated rabbit heart preparation. *J Am Coll Cardiol* 1983;2:465–473.
- [27] Mazgalev T, Dreifus LS, Bianchi J, Michelson EL. The mechanism of A–V junctional reentry: the role of the atrio-nodal junction. *Anat Rec* 1981;201:179–188.
- [28] Paes de Carvalho A. Cellular electrophysiology of the atrial specialized tissues. In: Paes de Carvalho A, de Mello WC, Hoffman BF, editors. *The Specialized Tissues of the Heart*. Amsterdam: Elsevier, 1961:113–115.
- [29] Watanabe Y, Dreifus LS. Inhomogeneous conduction in the A–V node. A model for reentry. *Am Heart J* 1965;70:505–514.
- [30] Moe GK, Cohen W, Vick RL. Experimentally induced paroxysmal A–V nodal tachycardia in the dog. A ‘case report’. *Am Heart J* 1963;65:87–92.

- [31] Lin F-Y, Lo H-M, Cheng J-J. Experimentally created atrioventricular node reentrant tachycardia in the dog: evidence of a brake system for nodal reentry in the anterior interatrial septum. *J Am Coll Cardiol* 1993;22:1541–1547.
- [32] Denes P, Wu D, Dhingra RC, Chuquimia R, Rosen KM. Demonstration of dual A–V nodal pathways in patients with paroxysmal supraventricular tachycardia. *Circulation* 1973;48:549–555.
- [33] Billette J. Atrioventricular nodal activation during periodic stimulation of the atrium. *Am J Physiol* 1987;252:H163–H177.
- [34] Moe GK, Preston JB, Burlington H. Physiologic evidence for a dual A–V transmission system. *Circ Res* 1956;4:357–375.
- [35] Simson MB, Spear J, Moore EN. The relationship between atrioventricular nodal refractoriness and the functional refractory period in the dog. *Circ Res* 1979;44:121–126.
- [36] Hoffman BF, Moore EN, Stuckey JH, Cranefield PF. Functional properties of the atrioventricular conduction system. *Circ Res* 1963;13:308–328.
- [37] Ferrier GR, Dresel PE. Relationship of the functional refractory period to conduction in the atrioventricular node. *Circ Res* 1974;35:204–214.
- [38] Loh P, de Bakker JMT. Unpublished observation (1997).
- [39] Loh P, de Bakker JMT, Hocini M, Thibault B, Janse MJ. High resolution mapping and dissection of the triangle of Koch in canine hearts: evidence for subatrial reentry during ventricular echoes. *PACE* 1997;20:1080 (Abstract).
- [40] McGuire MA, Janse MJ. New insights on anatomical location of components of the reentrant circuit and ablation therapy for atrioventricular junctional reentrant tachycardia. *Curr Opin Cardiol* 1995;10:3–8.
- [41] Josephson ME. *Clinical Electrophysiology: Techniques and Interpretation*. Philadelphia: Lea and Febiger, 1993.
- [42] Lewis T, Feil HS, Stroud WD. Observations upon flutter and fibrillation. Part II. The nature of auricular flutter. *Heart* 1920;7:191–245.
- [43] Puech P. *L'activité électrique circulaire normale et pathologique*. Paris: Masson and Cie., 1956.
- [44] Cosio FG. Endocardial mapping of atrial flutter. In: Touboul P, Waldo AL, editors. *Atrial Arrhythmias*. St. Louis: Mosby Year Book, 1990:229–240.
- [45] Hoffman BF. Experimental models of atrial flutter. In: Touboul P, Waldo AL, editors. *Atrial Arrhythmias*. St. Louis: Mosby Year Book, 1990:183–189.
- [46] Rosenbluth A, García Ramos J. Studies on flutter and fibrillation II. The influence of anatomical obstacles on experimental auricular flutter. *Am Heart J* 1947;33:677–684.
- [47] Kimura E, Kato K, Murao S, Ajisaka H, Koyama S, Omiya Z. Experimental studies on the mechanism of auricular flutter. *Tohoku J Exp Med* 1954;60:197–207.
- [48] Boineau JP, Schuessler RB, Mooney CR, et al. Natural and evoked atrial flutter due to circus movement in dogs. *Am J Cardiol* 1980;45:1167–1181.
- [49] Feld GF, Shahandeh-Rad F. Mechanisms of double potentials recorded during sustained atrial flutter in the canine right atrial crush-injury model. *Circulation* 1992;86:628–641.
- [50] Inoue H, Matsuo H, Takayanagi K, Murao S. Clinical and experimental studies of the effects of extrastimulation and rapid pacing on atrial flutter: evidence of macro reentry with an excitable gap. *Am J Cardiol* 1981;48:623–631.
- [51] Frame LH, Page RL, Hoffman BF. Atrial reentry around an anatomic barrier with a partially refractory excitable gap. A canine model of atrial flutter. *Circ Res* 1986;58:495–511.
- [52] Waldo AL. Mechanisms of atrial fibrillation, atrial flutter, and ectopic atrial tachycardia. A brief review. *Circulation* 1987;75:III37–III40. Suppl. III.
- [53] Boyden PA, Hoffman BF. The effects on atrial electrophysiology and structure of surgically induced right atrial enlargement in dogs. *Circ Res* 1981;49:1319–1331.
- [54] Pagé P, Plumb VJ, Okumura K, Waldo AL. A new model of atrial flutter. *J Am Coll Cardiol* 1986;8:872–879.
- [55] Allesie MA, Lammers WJEP, Bonke FIM, Hollen J. Intraatrial reentry as a mechanism for atrial flutter induced by acetylcholine in rapid pacing in the dog. *Circulation* 1984;70:123–135.
- [56] Mary-Rabine L, Mahaux V, Waleffe A, Kulbertus H. Atrial flutter: historical background. *J Cardiovasc Electrophysiol* 1997;8:353–358.
- [57] Garrey WE. The nature of fibrillatory contraction of the heart: its relation to tissue mass and form. *Am J Physiol* 1914;33:397–414.
- [58] Garrey WE. Auricular fibrillation. *Physiol Rev* 1914;4:215–250.
- [59] Allesie MA, Lammers WJEP, Bonke FIM, Hollen J. Experimental evaluation of Moe's multiple wavelet hypothesis of atrial fibrillation. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology and Arrhythmias*. New York: Grune and Stratton, 1985:265–275.
- [60] Moe GK, Abildskov JA. Atrial fibrillation as a self-sustaining mechanism independent of focal discharge. *Am Heart J* 1959;58:59–70.
- [61] Moe GK. On the multiple wavelet hypothesis of atrial fibrillation. *Arch Int Pharmacodyn Ther* 1962;140:183–188.
- [62] Schuessler RB, Grayson TM, Bromberg BI, Cox JL, Boineau JP. Cholinergically mediated tachyarrhythmias induced by a single extrastimulus in the isolated canine right atrium. *Circ Res* 1992;71:1254–1276.
- [63] Konings KTS, Kirchhof CJHJ, Smeets JRLM, Wellens HJJ, Penn OC, Allesie MA. High-density mapping of electrically induced atrial fibrillation in humans. *Circulation* 1994;89:1665–1680.
- [64] Cox JL, Canavan TE, Schuessler RB, Cain ME, Lindsay BD, Stone C. The surgical treatment of atrial fibrillation II: Intraoperative electrophysiologic mapping and description of the electrophysiologic basis of atrial flutter and fibrillation. *J Thorac Cardiovasc Surg* 1991;101:406–426.
- [65] Wijffels MCEF, Kirchhof CJHJ, Dorland R, Allesie MA. Atrial fibrillation begets atrial fibrillation: a study in awake chronically instrumented goats. *Circulation* 1995;92:1954–1968.
- [66] Attuel P, Leclercq JF, Coumel P. Atrial electrophysiological substrate remodeling after tachycardia in patients with and without atrial fibrillation. *PACE* 1995;18:804 pt. II.
- [67] Murgatroyd FD. In: AJ Camm, editor. *Nonpharmacological Management of Atrial Fibrillation*. Amonk NY: Futura Publishing, 1997.
- [68] Daoud EG, Bogun F, Goyal R, et al. Effect of atrial fibrillation on atrial refractoriness in humans. *Circulation* 1996;94:1600–1606.
- [69] Daoud EG, Knight BP, Weiss R, et al. Effect of verapamil and procainamide on atrial fibrillation-induced electrical remodeling in humans. *Circulation* 1997;96:1542–1550.
- [70] Goette A, Honeycutt C, Langberg JJ. Electrical remodeling in atrial fibrillation. Time course and mechanisms. *Circulation* 1996;94:2968–2974.
- [71] Coumel P, Attuel P, Lavallée JP, Flammang D, Leclercq JF, Slama R. Syndrome d'arythmie auriculaire d'origine vagale. *Arch Mal Coeur* 1978;71:645–656.
- [72] Murgatroyd FD, Camm AL. Atrial Arrhythmia. *Lancet* 1993;341:1317–1322.
- [73] Ravelli F, Allesie MA. Effects of atrial dilation on refractory period and vulnerability to atrial fibrillation in the isolated Langendorff-perfused rabbit heart. *Circulation* 1997;96:1689–1695.
- [74] Nazir SA, Lab MJ. Mechanoelectric feedback and atrial arrhythmias. *Cardiovasc Res* 1996;32:52–61.
- [75] Boyden PA, Tilley LP, Pham TD, Liu SK, Fenoglio Jr. JJ, Wit AL. Effects of left atrial enlargement on atrial transmembrane potentials and structure in dogs with mitral valve fibrosis. *Am J Cardiol* 1982;49:1896–1908.
- [76] Hordof AJ, Edie R, Malm JR, Hoffman BF, Rosen MR. Electrophysiologic properties and response to pharmacologic agents of fibers from diseased human atria. *Circulation* 1976;54:774–779.
- [77] Ten Eick RA, Singer DH. Electrophysiologic properties of diseased human atrium I. Low diastolic potential and altered cellular response to potassium. *Circ Res* 1979;44:545–557.

- [78] Le Heuzey JY, Boutjdir M, Gagey S, Lavergne T, Guize L. Cellular aspects of atrial vulnerability. In: Attuel P, Coumel P, Janse MJ, editors. *The Atrium in Health and Disease*. Mt Kisco NY: Futura Publishing, 1989:81–94.
- [79] Spach MS, Dolber PC. Relating extracellular potentials and their derivatives to anisotropic propagation at a microscopic level in human cardiac muscle: evidence for electrical uncoupling of side-to-side connections with increasing age. *Circ Res* 1986;58:356–371.
- [80] Moïse NS, Gilmour Jr. RF, Riccio ML. An animal model of spontaneous arrhythmic death. *J Cardiovasc Electrophysiol* 1997;8:98–103.
- [81] Gilmour Jr. RF, Moïse NS. Triggered activity as a mechanism for inherited ventricular arrhythmias in German shepherd dogs. *J Am Coll Cardiol* 1996;27:1526–1533.
- [82] Freeman LC, Pacioretti LM, Moïse NS, Kass RS, Gilmour Jr. RF. Decreased density of I_{to} in left ventricular myocytes from German shepherd dogs with inherited arrhythmias. *J Cardiovasc Electrophysiol* 1997;8:872–883.
- [83] Doe M, Ursell P, Lee RJ, Stilson C, Chin M, Moïse NS. Heterogeneous sympathetic innervation in German shepherd dogs with inherited ventricular arrhythmias and sudden death. *J Am Coll Cardiol* 1995;25:20A Abstract.
- [84] Schwartz PJ, Locati EH, Napolitano C, Priori, S. The long QT syndrome. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology: From Cell to Bedside*. Philadelphia: Saunders, 1995:788–811.
- [85] Kass RS, Davies MP. The roles of ion channels in an inherited heart disease: molecular genetics of the long QT syndrome. *Cardiovasc Res* 1996;32:433–454.
- [86] Leenhardt A, Glaser E, Burguera M, Nurnberg M, Maison-Blanch P, Coumel P. Short-coupled variant of torsade de pointes: a new electrocardiographic entity in the spectrum of ventricular tachyarrhythmias. *Circulation* 1994;89:206–215.
- [87] Eisenberg SJ, Scheinman MM, Dullet N. Polymorphous ventricular tachycardia in patients with normal cardiac function and QT interval. *Am J Cardiol* 1995;75:687–692.
- [88] Janse MJ, Wit AL. Electrophysiological mechanisms of ventricular arrhythmias resulting from myocardial ischaemia and infarction. *Physiol Rev* 1989;69:1049–1169.
- [89] Wit AL, Janse MJ. The ventricular arrhythmias of ischaemia and infarction. *Electrophysiological mechanisms*. Mount Kisco NY: Futura Publishing, 1993.
- [90] Janse MJ, Van Capelle FJL, Morsink H, et al. Flow of 'injury' current and patterns of excitation during early ventricular arrhythmias in acute regional myocardial ischaemia in isolated porcine and canine hearts. Evidence for 2 different arrhythmogenic mechanisms. *Circ Res* 1980;47:151–165.
- [91] Gray RA, Jalife J, Panfilov A, et al. Nonstationary vortexlike reentrant activity as a mechanism of polymorphic ventricular tachycardia in the isolated rabbit heart. *Circulation* 1995;91:2454–2459.
- [92] Myerburg RJ, Kessler KM, Castellanos A. Sudden cardiac death. Structure, function, and time-dependence of risk. *Circulation* 1992;85:2–10. Suppl.
- [93] Maseri A, Severi S, Marzullo P. Role of coronary arterial spasm in sudden coronary ischaemic death. *Ann NY Acad Sci* 1982;382:204–217.
- [94] Vermeulen JT, Tan HL, Rademaker H, et al. Electrophysiologic and extracellular ionic changes during acute ischaemia in failing and normal rabbit myocardium. *J Mol Cell Cardiol* 1996;28:123–131.
- [95] Winterton SJ, Turner MA, O'Gorman DJ, Flores NA, Sheridan DJ. Hypertrophy causes delayed conduction in human and guinea pig myocardium: accentuation during ischaemic perfusion. *Cardiovasc Res* 1994;28:47–54.
- [96] Watanabe I, Johnson TA, Buchanan J, Engle CL, Gettes LS. Effect of graded coronary flow reduction on ionic, electrical, and mechanical indexes of ischaemia in the pig. *Circulation* 1987;76:1127–1134.
- [97] Jenkins MG, Johnson TA, Engle C, Gettes LS. Metabolic protection by verapamil during graded coronary flow reduction independent of effect on baseline systolic function. Separation of mechanical and ionic markers of ischaemia. *Circulation* 1989;80:1870–1877.
- [98] Kaplinsky E, Ogawa S, Balke CW, Dreifus LS. Two periods of early ventricular arrhythmias in the canine acute myocardial infarction model. *Circulation* 1979;60:397–403.
- [99] Hill JL, Gettes LS. Effects of acute coronary artery occlusion on local myocardial extracellular K^+ activity in swine. *Circulation* 1980;61:768–778.
- [100] Wilde AAM, Asknes G. Myocardial potassium loss and cell depolarisation in ischaemia and hypoxia. *Cardiovasc Res* 1995;29:1–15.
- [101] Downar E, Janse MJ, Durrer D. The effect of acute coronary artery occlusion on subepicardial transmembrane potentials in the intact porcine heart. *Circulation* 1977;56:217–224.
- [102] Kléber AG, Janse MJ, van Capelle FJL, Durrer D. Mechanism and time course of S–T and T–Q segment changes during acute regional myocardial ischaemia in the pig heart determined by extracellular and intracellular recordings. *Circ Res* 1978;42:603–613.
- [103] Shaw RM, Rudy Y. Electrophysiologic effects of acute myocardial ischaemia. A mechanistic investigation of action potential conduction and conduction failure. *Circ Res* 1997;80:124–138.
- [104] Kléber AG, Janse MJ, Wilms-Schopman FJG, Wilde AAM, Coronel R. Changes in conduction velocity during acute ischaemia in ventricular myocardium of the isolated porcine heart. *Circ Res* 1986;73:189–198.
- [105] Smith WT, Fleet WF, Johnson TA, Engle CL, Cascio WE. The Ib phase of ventricular arrhythmias in ischaemic in situ porcine heart is related to changes in cell-to-cell electrical coupling. *Circulation* 1995;92:3051–3060.
- [106] Rudy Y, Quan W. A model study of the effects of the discrete cellular structure on electrical propagation in cardiac tissue. *Circ Res* 1987;61:815–823.
- [107] Janse MJ, Kléber AG. Electrophysiological changes and ventricular arrhythmias in the early phase of regional myocardial ischaemia. *Circ Res* 1981;49:1069–1081.
- [108] Coronel R, Fiolet JWT, Wilms-Schopman FJG. Distribution of extracellular potassium and its relation to electrophysiological changes during acute myocardial ischaemia in the isolated perfused porcine heart. *Circulation* 1988;77:1125–1138.
- [109] Coronel R, Fiolet JWT, Wilms-Schopman FJG, Ophthof T, Schaapherder AFM, Janse MJ. Distribution of extracellular potassium and electrophysiologic changes during two-stage coronary ligation in the isolated, perfused canine heart. *Circulation* 1989;80:165–177.
- [110] Coronel R. Distribution of extracellular potassium during myocardial ischaemia. Thesis, University of Amsterdam. Dordrecht: ICG Printing, 1988; p. 133.
- [111] Cascio WE, Yan G-X, Kléber AG. Early changes in extracellular potassium in ischaemic rabbit myocardium. The role of extracellular carbon dioxide accumulation and diffusion. *Circ Res* 1992;70:409–422.
- [112] Case RB, Felix A, Castellana FS. Rate of rise of myocardial PCO_2 during early myocardial ischemia in the dog. *Circ Res* 1979;45:324–330.
- [113] Wilde AAM, Escande D, Schumacher CA, et al. Potassium accumulation in the globally ischaemic mammalian heart. *Circ Res* 1990;67:835–843.
- [114] Cascio WE, Yan G-X, Kléber AG. Passive electrical properties, mechanical activity, and extracellular potassium in arterially perfused and ischaemic rabbit ventricular muscle. Effect of calcium entry blockade or hypocalcemia. *Circ Res* 1990;66:1461–1473.
- [115] Shattock MJ, Bers DM. Rat vs. rabbit ventricle: Ca flux and intracellular Na assessed by ion-selective microelectrodes. *Am J Physiol* 1989;256:C813–C822.

- [116] Baxter GF, Heads RJ, Yellon DM. Oxidative stress and the second window of protection after preconditioning. *Circulation* 1996;94:2992–2993. letter.
- [117] Speechly DM, Mocanu MM, Yellon DM. Protein kinase C. Its role in ischemic preconditioning in the rat. *Circ Res* 1994;75:586–590.
- [118] Boyden PA, Jeck CD. Ion channel function in disease. *Cardiovasc Res* 1995;29:312–3188.
- [119] Severs NJ. Gap junction alterations in the failing heart. *Eur Heart J* 1994;15:53–57. Suppl.
- [120] Peters NS. New insights into myocardial arrhythmogenesis: distribution of gap-junctional coupling in normal, ischaemic and hypertrophied human hearts. *Clin Sci (Colch)* 1996;90:447–452.
- [121] Spach MS, Boineau JP. Microfibrosis produces electrical load variations due to loss of side-to-side cell connections: a major mechanism of structural heart disease arrhythmias. *PACE* 1997;20:397–413.
- [122] Endo T, Ribeiro LGT, Cheung WM, Faria DB, Petranto M, Maroko PR. Relationship between the extent of the hypoperfused zone of the myocardium and the occurrence of ventricular fibrillation. *Am Heart J* 1983;105:915–920.
- [123] Austin M, Wenger TL, Harrell Jr. FE, Luzzi FA, Strauss HA. Effect of myocardium at risk on outcome after coronary artery occlusion and release. *Am J Physiol* 1982;243:H340–H345.
- [124] Bolli R, Fisher DJ, Entman ML. Factors that determine the occurrence of arrhythmias during acute myocardial ischaemia. *Am Heart J* 1986;111:261–270.
- [125] Rosen MR, Janse MJ, Myerburg RJ. Arrhythmias induced by coronary artery occlusion: What are the electrophysiological mechanisms? In: Hearse DJ, Manning AC, Janse MJ, editors. *Life-threatening Arrhythmias during Ischaemia and Infarction*. New York: Raven Press, 1987:11–47.
- [126] Meesmann W. Early arrhythmias and primary ventricular fibrillation after acute myocardial ischaemia in relation to preexisting coronary collaterals. In: Parratt JR, editor. *Early Arrhythmias Resulting from Myocardial Ischaemia*. London: Macmillan Press, 1982:93–112.
- [127] Meesmann W, Schulz FW, Schley G, Adolphsen R. Ueberlebensquote nach akutem experimentellen Koronarverschluss in Abhängigkeit von Spontankollateralen des Herzens. *Z ges exp Med* 1970;153:246–264.
- [128] Schaper W. Experimental infarcts and the microcirculation. In: Hearse DJ, Yellon DM, editors. *Therapeutic Approaches to Myocardial Infarct Size Limitation*. New York: Raven Press, 1984:79–90.
- [129] Trolese-Mongheal Y, Duchene-Marullaz P, Trolese J-F, Leinot M, Lamar J-C, Lacroix P. Sudden death and experimental acute myocardial infarction. *Am J Cardiol* 1985;56:677–681.
- [130] Hunt GB, Ross DL. Influence of infarct age on reproducibility of ventricular tachycardia induction in a canine model. *J Am Coll Cardiol* 1989;14:765–773.
- [131] Karagueuzian HS, Fenoglio Jr. JJ, Weiss MB, Wit AL. Prolonged ventricular tachycardia induced by premature stimulation of the canine heart after coronary artery occlusion and reperfusion. *Circ Res* 1979;44:833–846.
- [132] Kuck K-H, Costard A, Schlüter M, Kunze K-P. Significance of timing programmed electrical stimulation after acute myocardial infarction. *J Am Coll Cardiol* 1986;8:1279–1288.
- [133] Breithardt G, Borggrefe M, Haesten K. Role of programmed ventricular stimulation and noninvasive recording of ventricular late potentials for the identification of patients at risk of ventricular tachyarrhythmias after acute myocardial infarction. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology and Arrhythmias*. Orlando: Grune and Stratton, 1985:553–561.
- [134] Roy DE, Marchand E, Thérault P, Waters DD, Pelletier GB, Bourassa MG. Programmed ventricular stimulation in survivors of an acute myocardial infarction. *Circulation* 1985;72:487–494.
- [135] Shenassa M, Shenassa H. Prevention and termination of ventricular tachycardia and fibrillation by drugs and antitachycardia pacing. In: Allessie MA, Fromer M, editors. *Atrial and Ventricular Fibrillation: Mechanisms and Device Therapy*. Armonk NY: Futura Publishing, 1997:145–175.
- [136] Borggrefe M, Chen X, Hindricks G, et al. Catheter ablation of ventricular tachycardia in patients with coronary heart disease. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology: From Cell to Bedside*. Philadelphia: WB Saunders, 1995:1502–1517.
- [137] Lawrie GM, Pacifico A. Surgery for ventricular tachycardia. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology: From Cell to Bedside*. Philadelphia: WB Saunders, 1995:1547–1552.
- [138] Wilber DJ, Lynch JJ, Lucchesi BR. Postinfarction sudden death: significance at inducible ventricular tachycardia and infarct size in a conscious canine model. *Am Heart J* 1985;109:8–18.
- [139] Gardner PI, Ursell PC, Pham TD, Fenoglio Jr JJ, Wit AL. Experimental chronic ventricular tachycardia: anatomic and electrophysiologic substrates. In: Josephson ME, Wellens HJJ, editors. *Tachycardias: Mechanisms, Diagnosis, Treatment*. Philadelphia: Lea and Febiger, 1984:29–60.
- [140] Euler DE, Prood CE, Spear JF, Moore EN. The interruption of collateral blood flow to the ischaemic canine myocardium by embolization of a coronary artery with latex. Effects on conduction delay and arrhythmias. *Circ Res* 1981;49:97–108.
- [141] Wetstein L, Mark R, Kaplinsky E, et al. Histopathologic factors conducive to experimental ventricular tachycardia. *Surgery* 1985;98:532–538.
- [142] De Bakker JMT, Coronel R, Tasseron S, et al. Ventricular tachycardia in the infarcted, Langendorff-perfused human heart: role of the arrangement of surviving cardiac fibers. *J Am Coll Cardiol* 1990;15:1594–1607.
- [143] Zipes DP. Influence of myocardial ischaemia and infarction on autonomic innervation of the heart. *Circulation* 1990;82:1095–1105.
- [144] Gaide MS, Myerburg RJ, Kozlovskis PL, Bassett AL. Elevated sympathetic response of epicardium proximal to healed myocardial infarction. *Am J Physiol* 1983;245:H646–H652.
- [145] Herre JM, Wetstein L, Lin Y-L, Mills AS, Dae M, Thames MD. Effect of transmural versus nontransmural myocardial infarction on inducibility of ventricular arrhythmias during sympathetic stimulation in dogs. *J Am Coll Cardiol* 1988;11:414–421.
- [146] Billman GE, Schwartz PJ, Stone HL. Baroreceptor reflex control of heart rate: a predictor of sudden death. *Circulation* 1982;66:874–880.
- [147] Schwartz PJ, Billman GE, Stone HL. Autonomic mechanisms in ventricular fibrillation induced by myocardial ischaemia during exercise in dogs with healed myocardial infarction: An experimental model for sudden death. *Circulation* 1984;69:790–800.
- [148] Schwartz PJ, Vanoli E, Stramba-Badiale M, De Ferrari G, Billman GE, Foreman RD. Autonomic mechanisms and sudden death. New insights from analysis of baroreceptor reflexes in conscious dogs with and without a myocardial infarction. *Circulation* 1988;78:969–979.
- [149] La Rovere MT, Specchia G, Mortara A, Schwartz PJ. Baroreflex sensitivity, clinical correlates, and cardiovascular mortality among patients with a first myocardial infarction. A prospective study. *Circulation* 1988;78:816–824.
- [150] Farrell TG, Paul V, Cripps PV, et al. Baroreflex sensitivity and electrophysiological correlates in patients after acute myocardial infarction. *Circulation* 1991;83:945–952.
- [151] Legato MJ. The anatomic matrix as a factor in susceptibility to lethal arrhythmias in a canine model of sudden death. *J Mol Cell Cardiol* 1993;25:501–508.
- [152] Pressler ML. Passive electrical properties of cardiac tissue. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology: From Cell to Bedside*. Philadelphia: WB Saunders, 1990:108–122.
- [153] Pressler ML. Membrane properties of the cardiac conduction system: comparative aspects. *Proc Kon Ned Akad v Wet* 1990;93:477–487.

- [154] Zuanetti G, Corr PB. Antiarrhythmic efficacy of a new class III agent, UK-68,798, during chronic myocardial infarction: evaluation using three-dimensional mapping. *J Pharm Exp Ther* 1991;256:325–334.
- Task Force of the Working Group on Arrhythmias of the European Society of Cardiology The Sicilian Gambit. A new approach to the classification of antiarrhythmic drugs based on their actions on arrhythmogenic mechanisms. *Circulation* 1991;84:1831–1851.
- Task Force of the Working Group on Arrhythmias of the European Society of Cardiology The Sicilian Gambit. A new approach to the classification of antiarrhythmic drugs based on their actions on arrhythmogenic mechanisms. *Eur Heart J* 1991;12:1112–1131.
- [156] Spinelli W, Hoffman BF. Mechanisms of termination of reentrant atrial arrhythmias by class I and class III antiarrhythmic agents. *Circ Res* 1989;65:1565–1579.
- [157] Janse MJ, van der Steen ABM, van Dam RT, Durrer D. Refractory period of the dog's ventricular myocardium following sudden changes in frequency. *Circ Res* 1969;24:251–262.
- [158] Ypma JFAM. Adaptation of refractory period of rat ventricle to changes in heart rate. *Am J Physiol* 1972;223:894–897.
- [159] Janse MJ, Wilms-Schopman F, Opthof T. Mechanisms of antifibrillatory action of Org 7797 in regionally ischemic pig heart. *J Cardiovasc Pharmacol* 1990;15:633–643.
- [160] Vermeulen JT, McGuire MA, Opthof T, et al. Triggered activity and automaticity in ventricular trabeculae of failing human and rabbit hearts. *Cardiovasc Res* 1994;28:1547–1554.
- [161] Franz MR, Schaefer J, Schottler M, Seed WA, Noble MIM. Electrical and mechanical restitution of the human heart at different rates of stimulation. *Circ Res* 1983;53:815–822.
- [162] Detweiler DK, Patterson DF. The prevalence and types of cardiovascular disease in dogs. *Ann NY Acad Sci* 1965;127:481–516.
- [163] Gross DR. Animal models in cardiovascular research. 2nd edn. Dordrecht, Kluwer Academic Publishers, 1994.